

HEALTH CERTIFICATE FOR EXPORT OF BOVID SEMEN [Specifically Bovine (*Bos taurus*, *Bos indicus*, *Bison bison*), Water buffalo (*Bubalus bubalis*), Yak (*Bos grunniens*)] FROM FOOT-AND-MOUTH DISEASE-FREE MEMBER STATES OF THE EUROPEAN UNION TO THE UNITED STATES OF AMERICA

1. EU Member State of provenance and competent authority:	2. Health certificate No. This certificate is valid for 30 days.
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A. ORIGIN OF SEMEN

3. Approval number of the semen collection center _____

4. Name and address of the semen collection center:	5. Name and address of the consignor:
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4a. Name and address of the semen sexing facility, if applicable:

6. Country and place of loading:	7. Means of transport:
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B. DESTINATION OF SEMEN

8. Name and address of the consignee:

C. IDENTIFICATION OF SEMEN

9.1 Name of donor bull	9.2 Breed	9.3 Age	9.4 Identification Number	9.5 Number of straws	9.6 Date of collection	9.7 Collection code	9.8 Indicate one: sexed semen or non-sexed semen

10. Seal number of container(s):

D. HEALTH INFORMATION**Section A (to be signed by the Center Veterinarian)**

11. I, the undersigned Center Veterinarian of the described semen collection center, hereinafter "SCC," certify that:

11.1. All bovid animals in the above SCC were:

- 11.1.1 Established as residents only if admitted by a formal process of quarantine, observation, and testing as required by legislation in force, notably Annex B to Council Directive 88/407/EEC, as amended by Directive 2003/43/EC or in Regulation (EU) 2016/429/ Commission Delegated Regulation (EU) 2020/686;
- 11.1.2. Admitted to the SCC herd only after having been proven free of brucellosis, tuberculosis, bovine genital campylobacteriosis and trichomoniasis;
- 11.1.3. Admitted to the SCC herd only after having been proven free of viremia from persistent bovine viral diarrhea virus infection before entry into the SCC resident herd; and
- 11.1.4. Were tested annually for brucellosis, tuberculosis, bovine genital campylobacteriosis, and trichomoniasis.
- 11.1.5. The semen for export to the United States was either (SELECT ONE):
 - Collected prior to June 1, 2011; **OR**
 - The semen in the consignment was collected after June 1, 2011, from donors that were negative to two serum neutralization tests (using a 1:8 cutoff titer) for Schmallenberg virus, with the first performed within 30 days prior to collection, and the second between 28 and 60 days after collection. Tests were performed in a laboratory approved by the national Competent Authority.

11.2. In the SCC:

- 11.2.1. The SCC is certified as clinically free of paratuberculosis.
- 11.2.2. The herd was tested for brucellosis, tuberculosis, bovine genital campylobacteriosis, and trichomoniasis in its entirety with negative results at the most recent herd test prior to the period of semen collection for export to the United States of America (USA);
- 11.2.3. No clinical or other evidence of brucellosis, tuberculosis, bovine genital campylobacteriosis, trichomoniasis or leptospirosis was found since the most recent herd test and prior to the embarkation of semen to the United States;
- 11.2.4. There was no evidence to indicate that the donors have been affected with tuberculosis or brucellosis during the 12 months prior to the collection of semen for export to the United States;
- 11.2.5. There was no clinical evidence of infection by bovine viral diarrhea virus, bluetongue virus, enzootic hemorrhagic disease (EHD) or infectious bovine rhinotracheitis virus during the 60 days prior to and during the period of collection of semen for export to the United States; and
- 11.2.6. All bulls passed a testing program with negative results consistent with the World Organization for Animal Health (WOAH, formerly OIE) Terrestrial Animal Health Code (Article 4.5.5) or as outlined in Council Directive 88/407/EEC, as amended, in Regulation (EU) 2016/429 or in Regulation (EU) 2016/429 (Commission Delegated Regulation (EU) 2020/686); to detect persistent testicular bovine viral diarrhea virus infection prior to semen release.

11.3. Each donor bull for the semen described above:

- 11.3.1. Originated from a tuberculosis-free herd;
- 11.3.2. Was not corralled, pastured, or held with animals of lesser health status or under any restrictions which would make them ineligible to export semen to the United States during the 60 days prior to and during the period of collection of semen for export to the United States;
- 11.3.3. Was subjected with negative results to the test described in 11.4.1 to 11.4.4 within six months prior to or six months after collection of the semen described above;
- 11.3.4. Was subjected with negative results to the tests for bluetongue virus group (BTV) described in 11.4.6;
- 11.3.5. Was inspected on the date of semen collection and found to be free of clinical signs of diseases transmissible in semen.

- 11.4. Where reference is made to health tests, the following tests were carried out:
- 11.4.1. The cervical test for bovine tuberculosis described in the World Organization for Animal Health (WOAH, formerly OIE) Manual for Diagnostic Tests and Vaccines for Terrestrial Animals;
- 11.4.2. SELECT ONE for brucellosis testing:
- Buffered brucella antigen card test.
 - Rose bengal test.
 - Buffered plate agglutination test.
 - Indirect ELISA test for bovine brucellosis.
 - Competitive ELISA test for bovine brucellosis.
 - SELECT if this was performed: In accordance with the WOAH Manual of Diagnostic Tests and Vaccines for Terrestrial Animals, under the condition that samples that react positively were retested with negative results using a suitable confirmatory test such as the complement fixation test;
- 11.4.3. SELECT ONE for bovine genital campylobacteriosis (*Campylobacter fetus ssp. venerealis*) with negative testing results:
- A polymerase chain reaction (PCR) test.
 - Culture of preputial smegma.
- Note: The immunofluorescent antibody test may be used only as a screening test under the condition that samples that react positively must be retested using a suitable confirmatory test such as a PCR or culture of preputial smegma with negative results;
- 11.4.4. SELECT ONE for trichomoniasis (*Trichomonas fetus*) with negative results;
- PCR test.
 - Microscopic examination.
 - Culture of preputial smegma.
- 11.4.5. SELECT ONE for epizootic hemorrhagic disease (EHD).
- The animals reside in a Member State or region of the Member State where no cases of EHD have been reported within the previous 12 months, and where no serological evidence of EHD infection exists/existed for this period; OR
 - The following serotypes of EHD exist: _____ and animals were tested on two occasions by an agar gel immunodiffusion test (AGID) with negative results; OR
 - Testing was by competitive enzyme-linked immunosorbent assay (C-ELISA) AND a whole-blood PCR test for all the above-listed serotypes of EHD, with negative results using blood samples taken prior to, and not less than 21 days following collection of the semen (the two samples may not be taken more than 12 months apart). OR
 - Testing was by competitive enzyme-linked immunosorbent assay (C-ELISA) AND a virus neutralization test (VNT) for all the above-listed serotypes of EHD, with negative results using blood samples taken prior to, and not less than 21 days following collection of the semen (the two samples may not be taken more than 12 months apart).
- 11.4.6. The donor bull:
- SELECT ONE: Was tested for the bluetongue virus (BTV) group on blood serum performed prior to the first day of semen collection, at least every 60 days during the collection period, and between 21 and 60 days after semen collection, with negative results;
 - AGID test.
 - ELISA test.
 - OR (SELECT ONE)
 - Was tested with a whole blood PCR test for BTV group with one negative test at the beginning and conclusion of the collection period, and at least every 28 days during the period of semen collection.
 - Was tested with a whole-blood virus isolation test for BTV group with one negative test at the beginning and conclusion of the collection period, and at least every 7 days during the period of semen collection.

<p>11.5. The semen was collected and processed under my supervision and placed in individual ampules or straws which were permanently marked with the name of the donor, his registration number, or the collection code;</p> <p>11.6. Semen collection equipment which came into contact with bulls, or their secretions and excretions was thoroughly disinfected after each use, and good laboratory practices were followed during collection and processing of semen in order to minimize the possible introduction of microbial contamination;</p> <p>11.7. Antibiotics were added to the semen and semen extender in amounts and combinations consistent with the standards described in “Certified Semen Services (CSS) Minimum Requirements for Disease Control of Semen Produced for AI,” Appendix I, website: 202112136CSSMinReqJan2021-ENG_FINAL_v_4.pdf (naab-css.org).</p> <p>11.8. No biological products other than frozen semen or embryos qualified for shipment to the United States were present in the containers prior to use for export of semen to the United States;</p> <p>11.9. The storage and shipping containers are either new or cleaned and disinfected; and</p> <p>11.10. Only virgin liquid nitrogen was used to export semen to the United States.</p> <p>11.11. For sexed semen:</p> <p>11.11.1 The semen collected and processed under my supervision was shipped to the semen sexing facility within the Member State of collection under seal or was maintained under the oversight of a center or official veterinarian.</p> <p>11.11.2 Note: the semen sexing facility used to sex the semen is located in the Member State where the semen was collected. The facility has submitted a “Cleaning and Disinfection Standard Operating Protocol” reviewed and approved by the USDA, and is listed Approved EU and EFTA Bovine Semen Sexing Facilities Animal and Plant Health Inspection Service (usda.gov).</p>		
<p>12.1. Date and place</p>	<p>12.2. Name and qualification of the Center Veterinarian</p>	<p>12.3. Signature and stamp of the Center Veterinarian</p> <p>(The signature and stamp must be a different color than that of the printed template text.)</p>

